

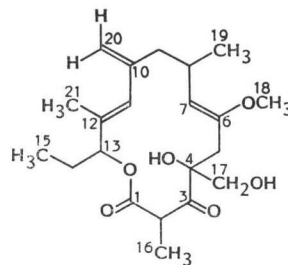
RUSTMICIN, A NEW MACROLIDE  
ANTIBIOTIC ACTIVE AGAINST  
WHEAT STEM RUST FUNGUS

Sir:

Stem rust (*Puccinia graminis* f. sp. *tritici*) is a serious wheat disease causing decrease of wheat production in many areas of the world<sup>1)</sup> and has been partially controlled so far by the use of resistant races<sup>2)</sup>, eradication of the alternate host, *i.e.* barberry<sup>3)</sup>, or by synthetic fungicides<sup>4-7)</sup>. We undertook the screening of new antibiotics active against wheat stem rust fungus using an inhibition assay of the germination of uredospores *in vitro* and pot test in green house. During the course of this screening, rustmicin (**I**) (formerly called P-59B1<sup>8)</sup>) was obtained from the cultured broth of *Micromonospora narashinoensis* 980-MC<sub>1</sub>. The present communication describes the isolation and structural elucidation of **I**.

Fermentation was carried out at 27°C with agitation of 400 rpm and aeration of 25 liters/minute for 4 days in two 50-liter jar fermentors each containing 25 liters of a medium consisting of soluble starch 2.5%, soybean meal 1.5%, dry yeast 0.2% and CaCO<sub>3</sub> 0.4% (pH 7.4). The activity against wheat stem rust fungus was monitored by an *in vitro* method, which assayed the germination of uredospores on the agar containing a sample 2 hours after inoculation at 20°C, throughout the purification. The active substance was isolated from the filtrate as described below. The cultured broth (*ca.* 47 liters) was harvested, adjusted to pH 7.0 with 2 N HCl and filtered. The filtrate was adsorbed on a Diaion HP20 column (5 liters) which was washed successively with H<sub>2</sub>O and 50% aq MeOH (10 liters each) and finally the active metabolite was eluted with MeOH (10 liters). The MeOH concentrate (0.5 liter) was partitioned between H<sub>2</sub>O and EtOAc three times. The combined organic layer (1 liter) was washed successively with 5% NaHCO<sub>3</sub>, 0.01 N HCl and H<sub>2</sub>O, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to afford a brownish oil (350 mg). This crude material was chromatographed over a Sephadex LH-20 column with MeOH and the active fractions were combined and evaporated *in vacuo* to dryness. The final purification of the antibiotic was achieved by reversed phase HPLC (Waters Radial PAK

Fig. 1. Structure of rustmicin (**I**).



8C<sub>18</sub>) eluting with 70% aq MeOH to yield 7.5 mg of **I** in a pure form.

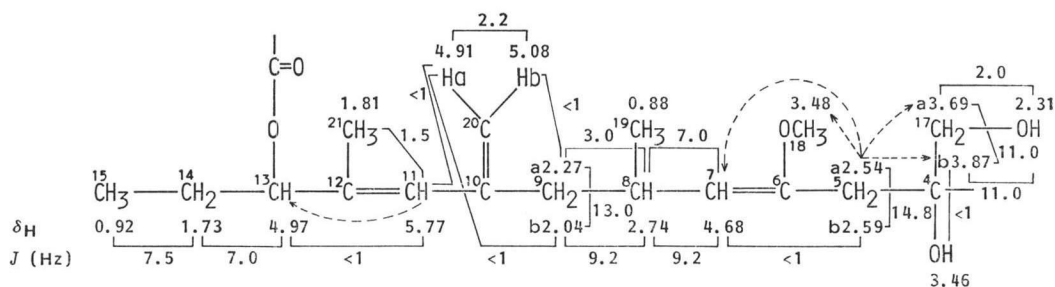
**I** is a neutral colorless oily material possessing the following physico-chemical properties: UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm ( $\epsilon$ ) 215 (9,500) and 240 (8,000); IR  $\nu_{\text{max}}^{\text{CHCl}_3}$  3600 (OH), 1730 and 1710 cm<sup>-1</sup> (COO and C=O); EI-MS  $m/z$  380 (M<sup>+</sup>). Its molecular formula was established to be C<sub>21</sub>H<sub>32</sub>O<sub>6</sub> by high resolution mass spectral data (found 380.2196; calcd 380.2200). The signals in the <sup>13</sup>C NMR spectrum of **I** taken in CDCl<sub>3</sub> accounted for 21 carbons and 30 nonexchangeable protons (see Table 1). Two protons at  $\delta_{\text{H}}$  2.31 (1H, dd,  $J=11.0$  and 2.0 Hz) and 3.46 (1H, br s) in the <sup>1</sup>H NMR spectrum of **I** revealed the presence of a primary and a tertiary alcohol, respectively.

In the <sup>13</sup>C NMR spectrum, the resonance at  $\delta_{\text{C}}$  169.2 was assigned to an ester carbonyl due to an IR absorption band at 1730 cm<sup>-1</sup>. The presence of a saturated ketone at  $\delta_{\text{C}}$  209.2 indicated that the UV absorption ( $\lambda_{\text{max}}$  240 nm) is due to a conjugated diene system. Thus, the functional groups in **I** are summarized as follows: CH<sub>3</sub> × 4, CH<sub>2</sub> × 3, CH × 2, CH<sub>3</sub>O × 1, CH<sub>2</sub>OH × 1, CH-O × 1, COH × 1, =CH<sub>2</sub> × 1, =CH × 2, =C × 3, COO- × 1 and C=O × 1. These data suggested the presence of one ring structure in **I**.

The structure of **I** was determined mainly based on spin decoupling and NOE experiments. The olefinic proton resonating at  $\delta_{\text{H}}$  5.77 (H-11; 1H, br s) showed long range couplings with a methyl at  $\delta_{\text{H}}$  1.81 and two protons at  $\delta_{\text{H}}$  4.91 and 4.97; irradiation of H-11 sharpened these three signals. The proton at  $\delta_{\text{H}}$  4.91 was coupled to the signal at  $\delta_{\text{H}}$  5.08 by a small coupling constant ( $J=2.2$  Hz). These two resonances were ascribed to an exomethylene whose <sup>13</sup>C resonance was observed at  $\delta_{\text{C}}$  116.9. The conjugated diene system in **I** proved to be located

Table 1.  $^{13}\text{C}$  NMR signals of I taken in  $\text{CDCl}_3$ .

| No. | Functional group | $\delta_{\text{C}}$ | No. | Functional group    | $\delta_{\text{C}}$ |
|-----|------------------|---------------------|-----|---------------------|---------------------|
| 1   | -COO-            | 169.2               | 12  | =C                  | 134.8               |
| 2   | -CH              | 51.1                | 13  | -CH-O               | 81.4                |
| 3   | -C=O             | 209.2               | 14  | -CH <sub>2</sub>    | 25.8                |
| 4   | -COH             | 81.8                | 15  | -CH <sub>3</sub>    | 9.9                 |
| 5   | -CH <sub>2</sub> | 33.9                | 16  | -CH <sub>3</sub>    | 13.9                |
| 6   | =C               | 148.2               | 17  | -CH <sub>2</sub> OH | 67.4                |
| 7   | =CH              | 124.1               | 18  | -OCH <sub>3</sub>   | 57.0                |
| 8   | -CH              | 30.7                | 19  | -CH <sub>3</sub>    | 21.0                |
| 9   | -CH <sub>2</sub> | 46.0                | 20  | =CH <sub>2</sub>    | 116.9               |
| 10  | =C               | 143.4               | 21  | -CH <sub>3</sub>    | 15.0                |
| 11  | =CH              | 129.3               |     |                     |                     |

Fig. 2. Partial structure of rustmicin (I).  
Dotted arrows indicate NOE observed.

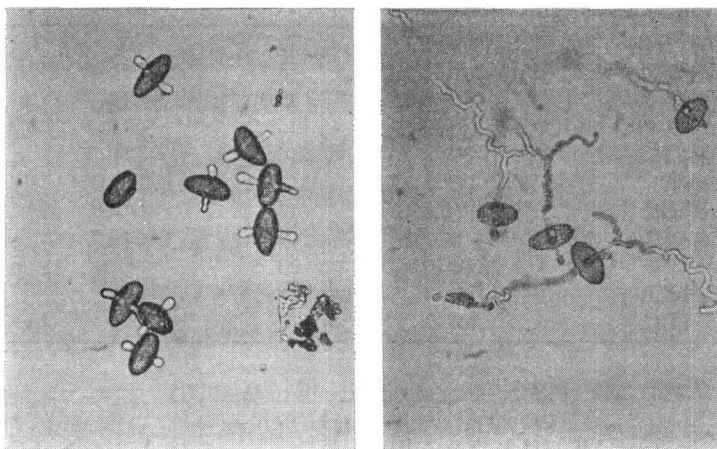
in the partial structure as shown in Fig. 2. Then, the  $^1\text{H}$  chemical shift of the only one oxymethine ( $\delta_{\text{H}}$  4.97,  $\delta_{\text{C}}$  81.4) mentioned above is responsible for the formation of an ester function. The connectivity from C-13 to C-15 and from C-7 to C-20 were straightforwardly revealed by the consecutive spin decoupling. The methine proton at  $\delta_{\text{H}}$  4.68 (H-7) was assigned to an olefinic proton based on a selective proton decoupling ( $\delta_{\text{C}}$  124.1). The allylic couplings between H-7 and methylene protons (H-5a and -5b) proved the connectivity from C-5 to C-7.

The connectivity from C-5 to C-15 was further extended by an NOE experiment; in addition to H-7, the irradiation of H-5a showed enhancements of the signals due to the methoxy group at  $\delta_{\text{H}}$  3.48 and the primary alcohol group at  $\delta_{\text{H}}$  3.69 and 3.87. This finding indicated either one of the methoxy ( $\delta_{\text{H}}$  3.48) or isolated hydroxymethyl was directly attached to C-6 and the other is combined to C-5 across the non-protonated carbon (C-4), *i.e.* the isolated carbonyl or *tert*-alcohol. The long range coupling observed between the hydroxymethyl proton at

$\delta_{\text{H}}$  3.87 and a broad singlet hydroxy proton at  $\delta_{\text{H}}$  3.46 enabled to determine the partial structure from C-5 to C-17. The double bond between C-6 and -7 proved to be in *Z* configuration by NOE enhancement observed with H-7 on the irradiation of H-5a as described above. Similarly, the geometry between C-11 and -12 was determined to *E* configuration by NOE enhancement with H-13 on the irradiation of H-11.

At this point, there remained two other partial structures,  $\text{CH}_3\text{CH}$  ( $\delta_{\text{H}}$  1.44, d and 3.73, q) and  $\text{C}=\text{O}$ , which can be accommodated in  $\alpha$ -ketolactone  $-\text{C}(\text{OH})\text{CH}(\text{CH}_3)\text{COCO}-$  or  $\beta$ -ketolactone  $-\text{C}(\text{OH})\text{COCH}(\text{CH}_3)\text{COO}-$ . Calculation of this methine attached to the ketone ( $\delta_{\text{H}}$  3.73) by the rule reported by CURPHEY<sup>9)</sup> indicated that the  $\beta$ -ketolactone ( $\delta_{\text{H}}$  3.65) was in good agreement with the observed value than the  $\alpha$ -ketolactone ( $\delta_{\text{H}}$  2.60). The methine substituted by two carbonyl and a methyl in pikromycin<sup>10)</sup> resonated at  $\delta_{\text{H}}$  4.30. This data supported the  $\beta$ -ketolactone system in I. Accordingly, the structure of rustmicin was elucidated as shown in Fig. 1.

Fig. 3. Inhibition of elongation of germ tube of *Puccinia graminis* by rustmicin (I): On the left is rustmicin (I) at 1  $\mu\text{g}/\text{ml}$  and, right, control.



The gross structure of rustmicin is similar to those of the 14-membered  $\beta$ -ketolactone antibiotics such as pikromycin<sup>10-12)</sup> and narbomycin<sup>13)</sup>, but different from nonglycosylated macrolides such as albocycline<sup>14,15)</sup>.

Rustmicin showed strong activity against wheat stem rust fungus *in vitro* and pot test in green house, MIC being 1 and 0.8  $\mu\text{g}/\text{ml}$ , respectively. Rustmicin inhibits the elongation of the germ tube accompanied with the swelling of tube tops *in vitro* (see Fig. 3). This unique antifungal effect is interesting as compared with the biological activities of the other macrolide antibiotics. Rustmicin exhibits antifungal but not significant antibacterial activity.

#### Acknowledgment

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#### References

- 1) ROELFS, A. P.: Estimated losses caused by rust in small grain cereals in the United States — 1918~76. U. S. Dept. Agric. Misc. Publ., 1363, 1978
- 2) ROELFS, A. P.; D. H. CASPER & D. L. LONG: Races of *Puccinia graminis* f. sp. *tritici* in the U.S.A. during 1978. Plant Dis. Repr. 63: 701~704, 1979
- 3) ROELFS, A. P.: Effects of barberry eradication on stem rust in the United States. Plant Dis. 66: 177~181, 1982
- 4) ROWELL, J. B.: Chemical control of the cereal rusts. Ann. Rev. Phytopathol. 6: 243~262, 1968
- 5) ROWELL, J. B.: Fungicidal management of pathogen populations. J. Environ. Qual. 1: 216~220, 1972
- 6) ROWELL, J. B.: Control of leaf rust on spring wheat by seed treatment with 4-*N*-butyl-1,2,4-triazole. Phytopathology 66: 1129~1134, 1976
- 7) ROWELL, J. B.: Control of stem rust on spring wheat by triadimefon and fenapanil. Plant Dis. 65: 235~236, 1981
- 8) ŌTAKE, N.; H. SETO, T. TAKATSU, A. SHIMAZU, T. SASAKI, T. SHOMURA, M. IWATA, T. WATANABE & T. ITO (Meiji Seika Kaisha): A new antibiotic P-59B1 and the process for the production thereof. Japan Kokai 85-6,197, Jan. 12, 1985
- 9) SILVERSTEIN, R. M.; G. C. BASSLER & T. C.

- MORRILL: Spectrometric Identification of Organic Compounds. 4th Ed., John Wiley and Sons, New York, p. 225, 1981
- 10) MUXFELDT, H.; S. SHRADER, P. HANSEN & H. BROCKMANN: The structure of pikromycin. J. Am. Chem. Soc. 90: 4748~4749, 1968
  - 11) TSAI, C.; J. J. STEZOWSKI & R. E. HUGHES: Crystal and molecular structure of kromycin. J. Am. Chem. Soc. 93: 7286~7290, 1971
  - 12) FURUHATA, K.; H. OGURA, Y. HARADA & Y. IITAKA: Stereochemistry of macrolides. II. Crystal structure of *p*-bromobenzoylpikromycin. Chem. Pharm. Bull. 25: 2385~2391, 1977
  - 13) PRELOG, V.; A. M. GOLD, G. TALBOT & A. ZAMOJSKI: Stoffwechselprodukte von Actinomyceten. 31. Über die Konstitution des Narbomycins. Helv. Chim. Acta 45: 4~21, 1962
  - 14) NAGAHAMA, N.; I. TAKAMORI & M. SUZUKI: Studies on an antibiotic, albocycline. IV. Catalytic hydrogenation and structure elucidation of albocycline. Chem. Pharm. Bull. 19: 655~659, 1971
  - 15) THOMAS, R. C. & C. G. CHIDESTER: Albocycline: Structure determination by X-ray crystallography. J. Antibiotics 35: 1658~1664, 1982